



Investigation into the 5-hydroxytryptamine-induced atropine-resistant neurogenic contraction of guinea-pig proximal colon

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1 The aim of this study was to characterize the receptors mediating the atropine-resistant neurogenic contraction to 5-hydroxytryptamine (5-HT) in the longitudinal muscle of the guinea-pig proximal colon and to determine the type of tachykinin receptors involved in the contractile response to 5-HT by the use of selective antagonists.

2 In the presence of atropine (0.3 μM), guanethidine (5 μM), hexamethonium (100 μM), ketanserin (0.1 μM) and indomethacin (3 μM), 5-HT (0.01–3 μM) produced concentration-dependent neurogenic contractions of colonic strips and at 0.3 μM produced a maximal effect ($\text{pEC}_{50} = 7.39 \pm 0.09$, $n = 18$). The 5-HT₄ receptor stimulant, 5-methoxytryptamine (5-MeOT, 0.03–10 μM) also produced neurogenic contractions with similar maximum effect to those of 5-HT ($\text{pEC}_{50} = 6.89 \pm 0.16$).

3 The 5-HT₄ receptor antagonist, DAU 6285 (3 μM) shifted the concentration-response curves to both 5-HT and 5-MeOT to the right without significant depression of the maximum, but the 5-HT₁/5-HT₂ receptor antagonist, metitepine (0.1 μM) and the 5-HT₃ receptor antagonist, ondansetron (0.3 μM) had no effect on the control curves to 5-HT and 5-MeOT.

4 The selective NK₁ receptor antagonist, FK 888 (1 μM) markedly attenuated the contractions to 5-HT and 5-MeOT. In contrast, the selective NK₂ receptor antagonist, SR 48968 (10 nM) and the selective NK₃ receptor antagonist, SR 142801 (10 nM) had no effect on the contractions to 5-HT and 5-MeOT.

5 These results indicate that the 5-HT-induced atropine-resistant neurogenic contraction of guinea-pig proximal colon is due to activation of 5-HT₄ receptors, presumably located on excitatory motor neurones, innervating the longitudinal muscle. The contraction evoked by activation of the 5-HT₄ receptors is mediated primarily via NK₁ receptors but not NK₂ or NK₃, suggesting that the 5-HT₄ receptor-mediated contraction is evoked indirectly via tachykinin release from tachykinin-releasing excitatory neurones.

Keywords: 5-Hydroxytryptamine (5-HT); 5-HT₄ receptors; atropine-resistant contraction; tachykinin receptors; proximal colon (guinea-pig)

Introduction

5-Hydroxytryptamine (5-HT) has been identified as a likely transmitter candidate for the regulation of intestinal motility (Furness & Costa, 1982). However, the precise role of 5-HT in the control of intestinal motility is still not clear.

In isolated preparations of the guinea-pig ileum and proximal colon, 5-HT produces contractions of the longitudinal muscle via an effect on cholinergic and non-cholinergic excitatory neurones (Costa & Furness, 1979). In the ileum, the 5-HT-evoked neurogenic contractions appear to be due to activation of two different receptor subtypes (5-HT₃ and 5-HT₄ receptors) located on the neurones (Buchheit *et al.*, 1985; Craig & Clarke, 1990; Eglen *et al.*, 1990). In the guinea-pig proximal and distal colon, atropine-sensitive contractions induced by 5-HT are mediated by the stimulation of 5-HT₃ and 5-HT₄ receptors, located on cholinergic neurones (Butler *et al.*, 1990; Elswood *et al.*, 1991; Wardle & Sanger, 1993), while the receptor subtypes responsible for atropine-resistant neurogenic contractions remain to be determined.

Substance P (SP) and the structurally related peptides, neurokinin A (NKA) and neurokinin B (NKB), belong to a family of biologically active peptides known as tachykinins, and their actions are mediated through three different types of receptors, NK₁, NK₂ and NK₃, which are preferentially activated by SP, NKA and NKB respectively (Mussap *et al.*, 1993, for review). Several lines of evidence converge to indicate tachykinins, SP and NKA, as the major excitatory transmitters

responsible for atropine-resistant transmission to the longitudinal muscle of the mammalian gut (Bartho & Holzer, 1985, for review).

Therefore, in this study we have examined the effects of selective 5-HT receptor antagonists in order to characterize the receptor(s) responsible for the 5-HT-induced atropine-resistant neurogenic contraction of the longitudinal muscle strips of the guinea-pig proximal colon. Additionally, we have investigated a role for tachykinin receptors in the contractile response to 5-HT using recently available, selective tachykinin receptor antagonists.

Methods

Tissue preparation

Male Dunkin Hartley guinea-pigs, weighing 250–500 g, were anaesthetized with halothane and bled. A 10 cm segment of the proximal colon starting 2 cm from the caecum was removed. Strips of mucosa-free longitudinal muscle from the proximal colon were prepared as described in a previous study (Kojima & Shimo, 1995). The strips were suspended in longitudinal direction under a 0.5 g load in 15 ml tissue baths filled with a modified Tyrode solution (mM: NaCl 136.8, KCl 2.7, CaCl₂ 1.8, MgCl₂ 1.05, NaH₂PO₄ 0.42, NaHCO₃ 11.9, glucose 5.56, ascorbic acid 0.12) (pH 7.4) at 37°C and bubbled with 5% CO₂-95% O₂. The modified Tyrode solution always contained atropine (0.3 μM) and guanethidine (5 μM) for the elimination of cholinergic and adrenergic responses and in-

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domethacin ($3 \mu\text{M}$), to minimize endogenous prostanoid biosynthesis in response to agonists. Hexamethonium ($100 \mu\text{M}$), was present to ensure that the responses recorded were the result of stimulation of postganglionic nerves. After setup, the strips were allowed to equilibrate for at least 60 min with renewal of the bathing solution every 15 min. Changes in mechanical activity of the tissue were recorded isotonically (isotonic transducer, Nihon Kohden, TD-112S; Nihon Kohden recticoder, RJG-4128).

Establishing concentration-response curves

In order to exclude an interference of the 5-HT_2 receptors mediating a contraction in the smooth muscle (Kojima, 1991), all the experiments designed to estimate 5-HT-induced NANC contractions were carried out in the presence of ketanserin ($0.1 \mu\text{M}$), a 5-HT_{2A} receptor antagonist. In the presence of ketanserin, noncumulative concentration-response curves for 5-HT or 5-methoxytryptamine (5-MeOT) within the range of $0.01\text{--}10 \mu\text{M}$ were established by adding increasing concentrations of the agonists to the tissue bath at intervals of at least 45 min. Each concentration was left in contact with the tissue for 1–2 min. In most cases, one of four preparations served as control and the three others were for the study of agonists in the presence of a set concentration of antagonist. Antagonists were allowed to equilibrate for 30–60 min prior to addition of agonists. The size of the contractions induced by agonists was expressed as the percentage of the response obtained with substance P (30 nM) in each preparation. Substance P was added at the first stage of the experimental protocol. The percent contractions were plotted as mean values to obtain log concentration-response curves.

Analysis of data

E_{max} (percentage of the effect of substance P, 30 nM) was the maximum contractile effect and pEC_{50} (or pEC_{30}) was the negative logarithm of the molar concentration of agonist causing 50% (or 30%) of the maximal contraction. pEC_{50} (or pEC_{30}) values were calculated according to Van Rossum (1963). Means \pm s.e. mean of n experiments are given throughout the paper. The significance of the difference between mean values was assessed by Student's unpaired t test, or by Dunnett's multicomparison test when appropriate. Results were considered significant if $P < 0.05$.

Drugs

The following drugs were used: atropine sulphate, hexamethonium chloride dihydrate (Wako, Osaka, Japan); guanethidine sulphate (Ciba, Basel, Switzerland); tetrodotoxin, ondansetron hydrochloride (Sankyo, Tokyo, Japan); substance P (Peptide Institute, Osaka, Japan); 5-HT creatine sulphate (Merck, Darmstadt, Germany); indomethacin, 5-MeOT hydrochloride (Sigma, St. Louis, MO, U.S.A.); DAU 6285 hydrochloride (endo-6-methoxy-8-methyl-8-azabicyclo [3.2.1] oct 3-yl-2, 3-dihydro-2-oxo-1H-benzimidazole-1 carboxylate hydrochloride, gift from Dr C.A. Rizzi, Boehringer Ingelheim, Milano, Italy); FK 888 ((2-(N-Me)indolil)-co-Hyp-Nal-NMe-Bzl, gift from Fujisawa Pharmaceutical Co., Ltd., Osaka, Japan); SR 48968 ((S)-N-methyl-N-[4-acetylamin-4-phenyl-piperidin-2-(3,4-dichlorophenyl)butyl] benzamide), SR 142801 ((S)-(N)-(1-(3-(1-benzoyl-3-(3,4-dichlorophenyl)piperidin-3-yl)propyl)-4-phenylpiperidin-4-yl)-N-methylacetamide, gift from Dr X. Emonds-Alt, Sanofi Recherche, Montpellier, France); ketanserin tartrate, metitepine maleate (Research Biochemicals Inc, Natick, U.S.A.).

All drugs were initially dissolved in saline with the following exceptions: indomethacin ($100 \mu\text{M}$) was dissolved in distilled water containing equimolar concentrations of Na_2CO_3 , SR 48968 ($100 \mu\text{M}$) and SR 142801 ($100 \mu\text{M}$) in 100% dimethylsulphoxide, 5-HT ($100 \mu\text{M}$) and 5-MeOT ($100 \mu\text{M}$) in saline containing ascorbic acid (0.12 mM). All subsequent di-

lutions of the drugs were made with saline, except for the 5-HT or 5-MeOT solutions, which always contained ascorbic acid (0.12 mM). The vehicles had no effects on 5-HT-induced contractions. The reported concentrations are the calculated final concentrations in the bath solution.

Results

Effects of 5-HT and 5-MeOT

In the presence of atropine ($0.3 \mu\text{M}$), guanethidine ($5 \mu\text{M}$), hexamethonium ($100 \mu\text{M}$), indomethacin ($3 \mu\text{M}$) and ketanserin ($0.1 \mu\text{M}$), 5-HT ($0.01\text{--}3 \mu\text{M}$) caused a phasic contraction in a concentration-dependent manner to a maximum effect at $0.3 \mu\text{M}$, amounting to $59.7 \pm 3.4\%$ of the effect of substance P (30 nM) and the pEC_{50} value was 7.39 ± 0.09 ($n = 18$) (Figures 1a and 2a). 5-Methoxytryptamine (5-MeOT, $0.03\text{--}10 \mu\text{M}$) mimicked the 5-HT-induced contraction with a pEC_{50} value of 6.89 ± 0.16 and a maximum effect at $3 \mu\text{M}$, amounting to $59.0 \pm 3.2\%$ ($n = 12$) of the effect of substance P (30 nM) (Figures 1b and 2b).

The contractions induced by 5-HT and 5-MeOT faded quickly in the continued presence of agonists (Figure 1). The responses to both agonists were almost abolished by tetrodotoxin (300 nM , Figure 2).

Effects of antagonists

Several antagonists with some degree of selectivity for the different receptor subtypes were tested against the 5-HT or 5-MeOT-induced contraction of guinea-pig proximal colon and these results are summarised in Table 1. None of the antagonists investigated had a significant influence on basal tone. Neither metitepine ($0.1 \mu\text{M}$), a $5\text{-HT}_1/5\text{-HT}_2$ receptor antago-

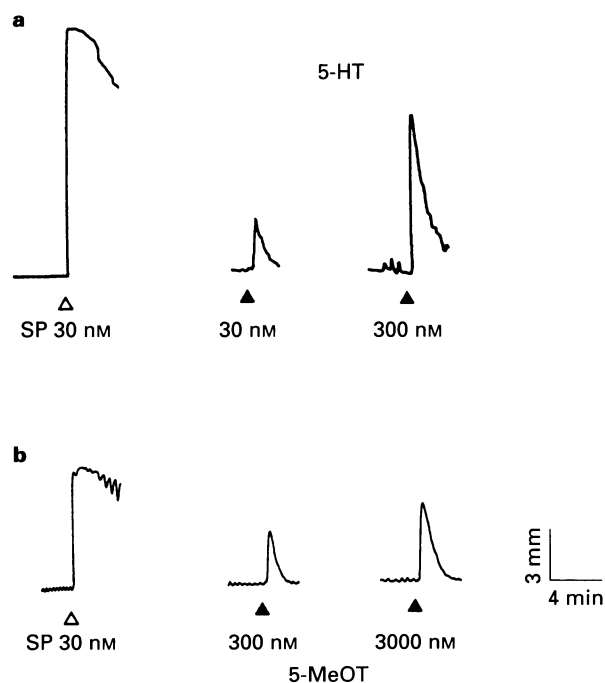


Figure 1 Typical tracings showing the contractile responses in the mucosa-free longitudinal muscle of guinea-pig isolated proximal colon to: (a) 5-hydroxytryptamine (5-HT); (b) 5-methoxytryptamine (5-MeOT). Arrows indicate the addition of agonists. Vertical calibration shows 3 mm shortening of the tissue, horizontal calibration shows 4 min. Atropine ($0.3 \mu\text{M}$), guanethidine ($5 \mu\text{M}$), hexamethonium ($100 \mu\text{M}$), indomethacin ($3 \mu\text{M}$) and ketanserin ($0.1 \mu\text{M}$) were present throughout the experiments.

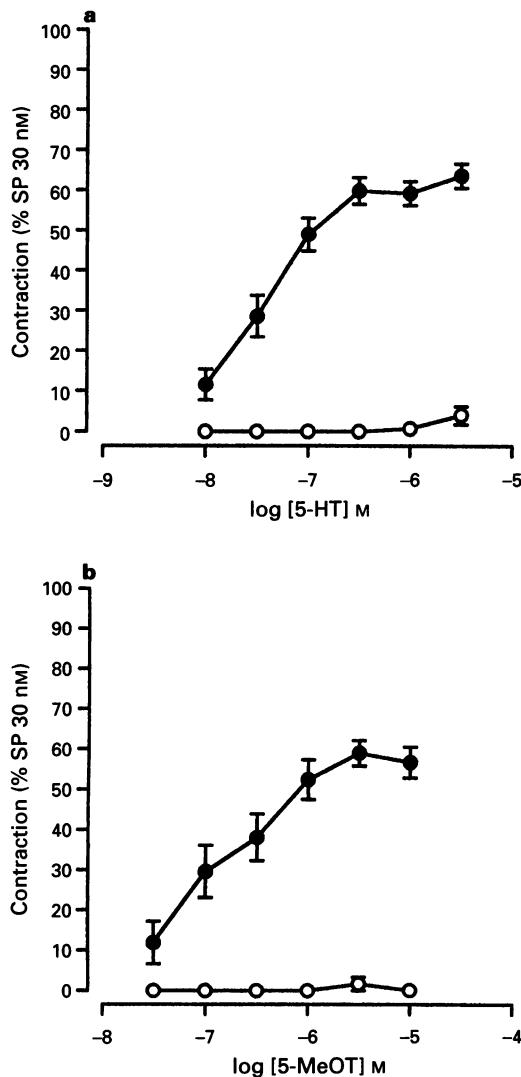


Figure 2 Concentration-response curves for 5-HT (a) and 5-MeOT (b)-induced contraction of guinea-pig proximal colon in the absence (●) and in the presence (○) of 300 nM tetrodotoxin. Each point represents the mean \pm s.e. mean of 6–18 preparations and is expressed as a percentage of the contractile response to 30 nM substance P. Atropine (0.3 μ M), guanethidine (5 μ M), hexamethonium (100 μ M), indomethacin (3 μ M) and ketanserin (0.1 μ M) were present throughout the experiments.

nist nor ondansetron (0.3 μ M), a 5-HT₃ receptor antagonist, caused any significant alteration in the concentration-response curves to 5-HT or 5-MeOT (Figure 3, Table 1, $P > 0.05$, Dunnet's test). DAU 6285 (3 μ M), a 5-HT₄ receptor antagonist displaced the concentration-response curves to both 5-HT and 5-MeOT to the right without significant depression of the maximum (Figure 3, Table 1). Furthermore, to evaluate the participation of tachykinin receptors in the contractile response to 5-HT, tachykinin receptor antagonists were studied. The contractile responses to both 5-HT and 5-MeOT were markedly depressed by FK 888 (1 μ M), a highly selective tachykinin NK₁ receptor antagonist (Figure 4, Table 1). The contraction evoked by substance P (3 nM, the amplitude of which was $72.9 \pm 5.4\%$ of 30 nM substance P ($n = 4$)-induced contraction) was also blocked by preincubation of the tissue with FK 888 (1 μ M) ($2.7 \pm 2.0\%$ of 30 nM substance P, $n = 4$, $P < 0.001$). Neither SR 48968 (10 nM), a highly selective tachykinin NK₂ receptor antagonist nor SR 142801 (10 nM), a highly selective tachykinin NK₃ receptor antagonist, affected the concentration-response curves to 5-HT or 5-MeOT (Figure 4, Table 1, $P > 0.05$, Dunnet's test).

Discussion

The present study has shown that 5-HT produces concentration-dependent and reproducible contractions of guinea-pig proximal colon in the presence of atropine, guanethidine, hexamethonium, ketanserin and indomethacin. Since tetrodotoxin almost totally abolished the contractions, this response is due to stimulation of postganglionic neurones, and is not due to activation of mechanisms involving the release of acetylcholine, noradrenaline or prostanooids.

The aims of this study were two fold. First, to characterize the receptor mediating the neurogenic contractions to 5-HT in the guinea-pig proximal colon and secondly to determine the type of tachykinin receptors involved in the contractile responses to 5-HT.

According to the current classification, four main types of functional 5-HT receptors, termed 5-HT₁, 5-HT₂, 5-HT₃, 5-HT₄ can be distinguished (Hoyer *et al.*, 1994). Studies with 5-HT receptor antagonists have given some insight into the types of 5-HT receptor responsible for the 5-HT-induced contraction. Metitepine, a 5-HT₁/5-HT₂ receptor antagonist, failed to block the 5-HT-induced contraction, suggesting that neither the 5-HT₁ nor 5-HT₂ receptor class makes a contribution to the contractile response to 5-HT.

It has repeatedly been shown that 5-HT₃ and 5-HT₄ receptors are on excitatory motor neurones which supply the longitudinal muscle of the guinea-pig ileum, proximal colon and distal colon (Eglen *et al.*, 1990; Butler *et al.*, 1990; Wardle & Sanger, 1994). However, in the present study using long-

Table 1 The potencies of 5-hydroxytryptamine (5-HT) and 5-methoxytryptamine (5-MeOT) in guinea-pig proximal colon in the absence and presence of various antagonists

Antagonists	5-HT		5-MeOT	
	pEC_{50} (or pEC_{30})	E_{max} (%)	pEC_{50} (or pEC_{30})	E_{max} (%)
Control	7.39 ± 0.09 (7.62 ± 0.09) ^a	59.7 ± 3.4	6.89 ± 0.16 (7.18 ± 0.16) ^a	59.0 ± 3.2
Metitepine (0.1 μ M)	7.23 ± 0.15	65.5 ± 7.1	6.86 ± 0.12	59.4 ± 4.3
Ondansetron (0.3 μ M)	7.28 ± 0.04	61.6 ± 6.0	6.74 ± 0.12	59.4 ± 6.3
DAU 6285 (3 μ M)	6.53 ± 0.21 ^{a*}	50.8 ± 8.8	6.07 ± 0.23 ^{a*}	39.6 ± 9.4
FK 888 (1 μ M)		4.3 ± 1.6 ^{**}		14.5 ± 7.2 ^{**}
SR 48968 (10 nM)	7.37 ± 0.08	49.6 ± 1.9	6.76 ± 0.11	65.0 ± 6.8
SR 142801 (10 nM)	7.15 ± 0.21	64.3 ± 5.2	6.65 ± 0.19	52.1 ± 5.7

Listed are pEC_{50} values (or pEC_{30}) and E_{max} values (percentage of the effect of substance P, 30 nM). The data are means \pm s.e. mean from 6–18 preparations. Atropine (0.3 μ M), guanethidine (5 μ M), hexamethonium (100 μ M), indomethacin (3 μ M) and ketanserin (0.1 μ M) were present throughout experiments. * $P < 0.001$ versus control pEC_{30} (Student's *t* test). ** $P < 0.01$ versus control E_{max} (Dunnet's test).

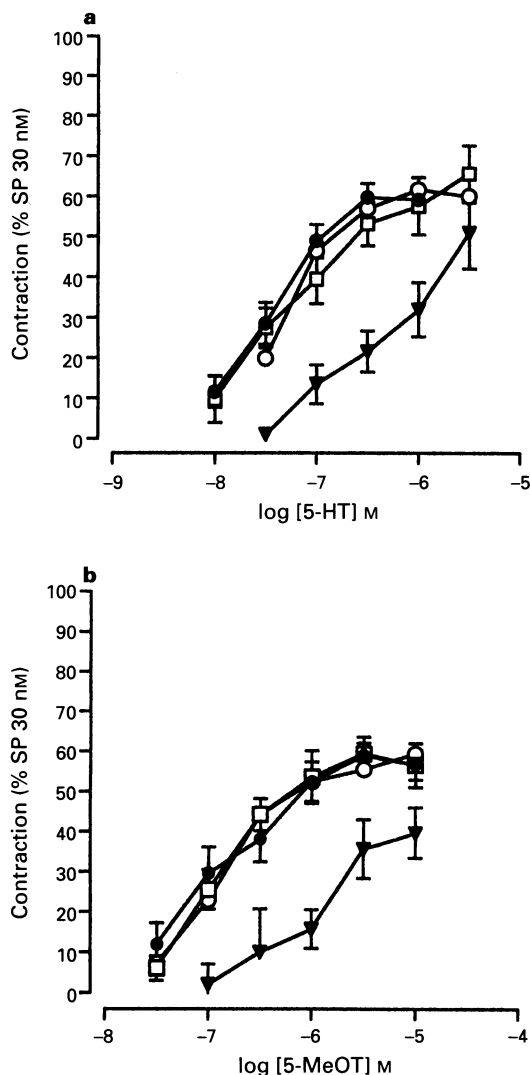


Figure 3 Concentration-response curves for 5-HT (a) and 5-MeOT (b)-induced contraction of guinea-pig proximal colon in the absence (●) and in the presence of 0.1 μM metitepine (□); 0.3 μM ondansetron (○); 3 μM DAU 6285 (▼). Each point represents the mean ± s.e. mean of 6–18 preparations and is expressed as a percentage of the contractile response to 30 nM substance P.

itudinal strips of proximal colon, we have found no evidence for the involvement of 5-HT₃ receptors in mediating contractile responses to 5-HT because ondansetron, a 5-HT₃ receptor antagonist, failed to modify the 5-HT concentration-response curve. There are two possible reasons for this discrepancy. Firstly, 5-HT₃ receptor-mediated responses only become significant from 1 μM and above (Eglen *et al.*, 1990; Butler *et al.*, 1990; Wardle & Sanger, 1994), whereas we tested 5-HT only up to 3 μM and 5-MeOT is not an agonist at 5-HT₃ receptors. Secondly, differences in effective 5-HT₃ receptor reserves (leading to tachykinin release) in different regions of the intestine may be involved, since the 5-HT₃ receptor-mediated contraction of guinea-pig ileum was inhibited by D-Pro⁴, D-Trp^{7,9} substance P (4-11), a tachykinin receptor antagonist (Buchheit *et al.*, 1985), whereas spantide, a tachykinin receptor antagonist caused only a slight inhibition of 5-HT₃ receptor-mediated contraction of guinea-pig distal colon (Woollard *et al.*, 1994). On the other hand, in the present study, DAU 6285, a 5-HT₄ receptor antagonist was effective in antagonizing the contractile effect of 5-HT, suggesting an involvement of the 5-HT₄ receptor subtype in the contractile response to 5-HT. Additional support for this contention is provided by results of experiments where the effects of the 5-HT₄ receptor stimulant, 5-MeOT were examined. We found that 5-MeOT mimicked

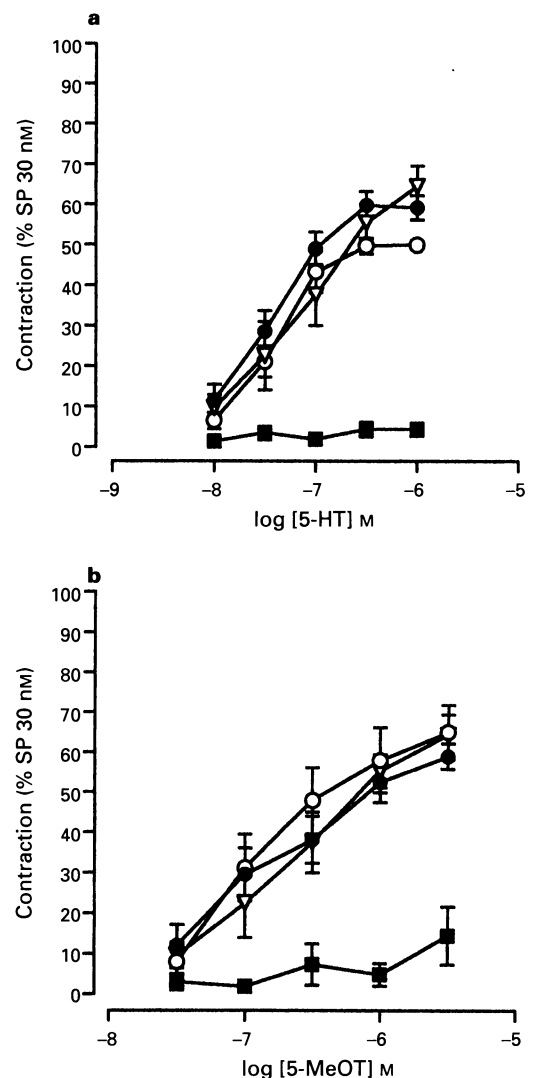


Figure 4 Concentration-response curves for 5-HT (a) and 5-MeOT (b)-induced contraction of guinea-pig proximal colon in the absence (●) and in the presence of 1 μM FK 888 (■); 10 nM SR 48968 (○); 10 nM SR 142801 (▼). Each point represents the mean ± s.e. mean of 6–18 preparations and is expressed as a percentage of the contractile response to 30 nM substance P.

the 5-HT-induced contraction and that the 5-MeOT-induced contractions were antagonized by DAU 6285. Thus, these results suggest that the 5-HT-induced atropine-resistant contraction is due to activation of 5-HT₄ receptors. We have also examined the effects of selective NK₁ and NK₂ receptor antagonists in order to gain information on the nature of NK receptor types involved in the atropine-resistant contraction induced by 5-HT. FK 888 has been characterized as a highly selective NK₁ receptor antagonist ($pK_B = 7.13$), while SR 48968 is a potent and highly selective NK₂ receptor antagonist ($pK_B = 9.41$) in the guinea-pig proximal colon (Maggi *et al.*, 1994). The present results show that the NK₁, but not the NK₂ receptor antagonist markedly attenuated the contractions induced by 5-HT or 5-MeOT, suggesting that NK₁ receptors are the primary tachykinin receptors which mediate the contractions upon 5-HT₄ receptor activation. This result is also compatible with previous work indicating a major role for NK₁ receptors in mediating non-cholinergic excitatory junction potentials and contractions evoked by electrical field stimulation in the guinea-pig proximal colon (Zagorodnyuk *et al.*, 1993) or in mediating substance P-induced contractions of guinea-pig proximal colon (Briejer *et al.*, 1993). Furthermore, we have made use of the recently described NK₃ receptor antagonist, SR 142801 (Emonds-Alt *et al.*, 1995) to define a role

for NK₃ receptors because previous work indicated that the activation of NK₃ receptors on myenteric plexus neurones results in acetylcholine and tachykinin release (Guard & Watson, 1987). However, the possibility that NK₃ receptors play a role in the contractile response to 5-HT is unlikely, because 10 nM SR 142801, a potent and highly selective NK₃ receptor antagonist ($pA_2 = 9.15$ in the guinea-pig ileum) (Emonds-Alt *et al.*, 1995) had no effect upon the contractile responses to 5-HT or 5-MeOT. Thus, these results suggest that the contractile responses to 5-HT₄ receptor activation are evoked indirectly via tachykinin release from tachykinin-releasing neurones. The exact nature of the tachykinin responsible for the 5-HT-induced atropine-resistant contraction has not yet been determined. However, since substance P and neurokinin A have been shown to be co-localized in myenteric plexus neurones (Deacon *et al.*, 1987), and further both these tachykinins can stimulate NK₁ receptors with differing affinity and efficacy (Mussap *et al.*, 1993), both these tachykinins may be involved in the atropine-resistant contraction. On the other hand, co-existence of acetylcholine with tachykinins has been repeatedly described in excitatory enteric neurones (Furness *et al.*, 1992), and it is thus to be expected that, if 5-HT₄ receptor stimulation leads to acetylcholine release, the release of the tachykinins is also enhanced. Indeed, the present study is congruent with the idea that the 5-HT₄ receptor-mediated contraction is due to the co-release of acetylcholine and tachykinins from the same population of enteric neurones, but coexistence of 5-HT with tachykinins has also been described in myenteric neurones of guinea-pig colon (Legay *et al.*, 1984). Thus, it is also possible that 5-HT₄ receptor activation causes the release of tachykinins from a separate population of non-cholinergic neurones.

In conclusion, these results indicate that the 5-HT-induced atropine-resistant contraction is mediated by the stimulation of 5-HT₄ receptors, located on excitatory motor neurones which supply the longitudinal muscle of guinea-pig proximal colon. The contraction evoked by activation of the 5-HT₄ receptors is mediated primarily via NK₁ receptors but not NK₂ or NK₃ receptors, suggesting that the 5-HT₄ receptor-mediated contraction is evoked via endogenously released tachykinins. Thus, our data support our previous hypothesis indicating the presence of prejunctional 5-HT₄ receptors localized on tachykinin-releasing neurones in the guinea-pig proximal colon (Kojima & Shimo, 1995). However, the exact physiological importance of the 5-HT₄ receptors described still needs to be determined.

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